

ACP₁ – ADA₁ Interaction and Human Health

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Abstract:

Background: Biochemical and epidemiological evidence suggest that ACP₁ and ADA₁ genetic polymorphisms interact with significant phenotypic effects at the clinical level. We have reviewed the observations collected by our group over twenty years on the interaction between these polymorphisms in order to elucidate their impact on human health.

Methods: ACP₁ and ADA₁ phenotypes from 3297 subjects were determined by starch gel electrophoresis on RBC hemolysates.

Results: Birth weights are higher in newborns with the ACP₁*A/*A and *B/*A genotypes carrying the ADA₁*2 than in those with other joint genotypes. During the pre-reproductive and early reproductive stages of extrauterine life, individuals carrying these genotypes are more susceptible to diseases involving the immune system. ACP₁*A/*A and *B/*A genotypes carrying the ADA₁*2 allele are protected against common diseases occurring during the last stage of life.

Conclusions: The joint ACP₁-ADA₁ system has an important role in human health and development.

Keywords: ACP₁; ADA₁; human health; genetic interaction

Introduction

Biochemical and epidemiological evidence suggest that Acid Phosphatase locus 1 (ACP₁) and Adenosine Deaminase locus 1 (ADA₁) genetic polymorphisms interact with significant phenotypic effects at the clinical level. In the present paper we review the observations collected by our group in the last twenty years in order to elucidate their roles on human health [1-7].

Genetic polymorphism of ACP₁

ACP₁ is a polymorphic enzyme controlled by a gene on chromosome 2. In European population there are 3 common codominant alleles: ACP₁*A, ACP₁*B and ACP₁*C. ACP₁ functions as a phosphatase and these three distinct alleles exhibit markedly different enzymatic activities. The total enzymatic activity for the six ACP₁ phenotypes measured with p-nitrophenylphosphate as a substrate, is in the order *A/*A<*A/*B<(*B/*B<A/*C)<*B/*C<*C/*C.

Two biological substrates have been suggested for ACP₁: flavin-mononucleotide (FMN) and phosphorylated protein-tyrosine residues. Catalysing the conversion of FMN to riboflavin, ACP₁ may regulate the cellular concentration of Flavin Adenine Dinucleotide (FAD), flavo-enzyme activity and energy metabolism. As a protein phosphotyrosine phosphatase, ACP₁ may have an important role in modulating the glycolytic rate through the control of insulin receptor activity and of band 3 protein phosphorylation status [8].

ACP₁ dephosphorylates a negative phosphorylation site in Zeta-chain-associated protein kinase 70 (ZAP 70) in T cells [9]. This event leads to increased activation of the kinase and enhanced signalling from T cell antigen receptor.

ACP₁ negatively modulates insulin signal transduction [10] and is able to dephosphorylate the Adipocyte Lipid Binding Protein (ALBP) in vitro [11]. ALBP belongs to a family of lipid binding proteins present in various isoforms in many human tissues. In adipose tissue ALBP is phosphorylated on Tyr19 after insulin stimulation thereby impairing its fatty acid binding ability. In the adipose tissue the dual activities of ACP₁ (on insulin receptor transduction and ALBP dephosphorylation) could partly compensate for each other.

Genetic polymorphism of ADA₁

ADA₁ is a polymorphic enzyme present in all mammalian tissues. It is controlled by a gene with two codominant alleles ADA₁*1 and ADA₁*2 located on the long arm of chromosome 20. Similar to ACP₁ the polymorphism of this enzyme exhibits different enzymatic activities between phenotypes in the order ADA₁*1/*1>ADA₁*2/*1>ADA₁*2/*2. ADA₁ catalyses the irreversible deamination of adenosine to inosine thereby contributing to the regulation of intracellular and extracellular concentration of adenosine. Experimental studies have shown that adenosine, acting via the A1R, increases insulin sensitivity in isolated adipocytes [12-14] and



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decreases insulin sensitivity in isolated muscle fibres [15,16].

ADA was thought to be an exclusively cytosolic enzyme until the recent demonstration that it is present on the surface of erythrocytes and platelets. Subsequently, it has been demonstrated that ADA is present on the surface of many cell types including lymphocytes and neurons, where it can act as an ectoenzyme [17,18]. The functional role of ecto-ADA is related not only to degradation of extracellular adenosine [19], but also to transduction signals through its interaction with either CD26 or Adenosine 1 Receptor (A1R), two ADA anchoring proteins. In this manner ADA acting as a costimulatory molecule integrates multiple signalling events in different cell types [20]. Adenosine, a purine nucleoside present in plasma and other extracellular fluids is an important local hormone (like prostaglandins) that regulates blood flow, neurotransmission, physiology of smooth muscle and platelet aggregation. Current interest has been focused on a wide variety of effects produced by adenosine via activation of cell surface adenosine receptors [21,22]. Recent studies indicates that adenosine counteracts insulin action in the liver by activating A2B receptors [23] while also facilitating insulin action in adipocytes.

Adenosine Deaminase Complex Protein (ADPC) is identical with CD26 [24], a T cell activating antigen and with a glycoprotein present in epithelial cells of various tissues. Recent data suggest that ADA₁ and CD26 are co-localized on the T cell surface but not inside cells. Cells expressing ADA₁ and CD26 on the surface are much more resistant to the inhibitory effects of adenosine. These data suggest that ADA₁ on the cell surface is involved in an important immunoregulatory mechanism by which released ADA₁ binds to the cell surface of CD26, and this

We studied the following samples:

Children with Type 1 diabetes
 Women with Type 1 diabetes
 Women with repeated spontaneous abortion
 Allergic women with endometriosis
 Crohn's disease
 Type 2 diabetes
 Subjects with Cardiovascular Diseases
 Blood donors (2 samples)
 Healthy newborn infants
 Children from the Central Area of Sardinia

ACP₁ and ADA₁ phenotype were determined as previously described [1-7]. Test of independence was carried by Chi-square and differences between means evaluated by Student-t. Three way contingency table to test interaction among variables was carried out by

complex is capable of reducing the local concentration of adenosine [19].

The interaction between ACP₁ and ADA₁

ACP₁ is modulated by various substances including purines [25] and its activity is influenced by ADA₁ genotype: in carriers of ADA₁*2 allele the activity is generally lower than in homozygotes for the ADA₁*1 allele [2]. There is also an interaction between ADA₁ and ACP₁ genotypes concerning their effects on ACP₁ activity. ACP₁*A/*A carrying the ADA₁*2 allele shows a 37% lower activity than ACP₁*A/*A with the ADA₁*1/*1 genotype; ACP₁*A/*B carrying the ADA₁*2 allele shows a 13% lower activity than ACP₁*A/*B with the ADA₁*1/*1 type, whereas for other ACP₁ types no significant differences have been observed between carriers of ADA₁*2 and subjects with ADA₁*1/*1 genotype. As a result, a very low ACP₁ activity is observed in individuals having ACP₁*A/*A genotype and carrying ADA₁*2 allele. Overall in *A/*A and *B/*A phenotypes carrying the ADA₁*2 allele there is a mean decrease of 18.5% of ACP₁ enzymatic activity. Therefore, effects related to low ACP₁ activity (i.e. *A/*A and *B/*A genotypes) could be modified in individuals carrying the ADA₁*2 allele.

Adenosine and inosine show modulating effects on ACP₁. The activation by inosine is much higher than by adenosine. Since the deamination rate constant of adenosine to inosine associated with ADA₁*2/*1 phenotype is slower than that associated with ADA₁*1/*1, the interaction of ADA₁ on ACP₁ activity may be explained by a lower intracellular concentration of inosine in ADA₁*2 carriers.

Material and Methods

A total of 1913 of subjects from the population of Central Continental Italy and a total of 1384 subjects from the population of Central Sardinia were examined.

286 subjects (Ref 6)
 107 subjects (Ref 7)
 170 subjects (Ref 4)
 38 subjects (Ref 5)
 70 subjects (unpublished)
 273 subjects (Ref 3)
 339 subjects (unpublished)
 290 subjects (unpublished)
 340 subjects (Ref 1)
 1384 subjects (Ref 7)

log-linear model. All analyses were carried out by commercial software (SPSS).

All subjects gave written informed consent to participate to the study that was conform with The Code of Ethics of the World Medical Association.

Results

Table 2 shows the proportion of low activity ACP₁ genotypes (*A/*A and *A/*B) in ADA₁*1/*1 subjects and in carriers of ADA₁*2 allele. Five classes of subjects with immunological disease and two samples of blood donors are reported. In all samples of immunological diseases the proportion of low activity ACP₁ genotypes is higher in carriers of ADA₁*2 allele than in ADA₁*1/*1 genotype. Overall the ODDS ratio is 1.967 (95% C.I. 1.278-3.030). The pattern of association is different from that observed in blood donors in whom the proportion of low activity ACP₁ genotypes is higher in ADA₁*1/*1 genotype than in carriers of ADA₁*2 allele. There is a statistically significant interaction suggesting that ADA₁ genotype modifies the pattern of association between ACP₁ and the diseases considered: low activity ADA₁ could lower *A/*A and *A/*B activities and in turn ZAP 70 activity resulting in a weakening of T cell receptor signaling. An additive effect seems unlikely since a high concentration of adenosine, due to low activity of ADA₁ genotypes carrying the ADA₁*2 allele, would contribute to strengthen T cell activation (see table 1) with an effect opposite to that observed in association with low ACP₁ activity.

Table 3 shows the proportion of *A/*A and *A/*B genotypes carrying the ADA₁*2 allele in subjects with immunological diseases and in blood donors. These joint genotypes are much more frequent in the immunological diseases than in blood donors (p=0.0013; O.R. 2.31 95% C.I. 1.465-5.910) suggesting that these genotypes predispose to the diseases considered.

In fig 1 we compared patients with immunological diseases to blood donors and consecutive health newborns from the same population. The proportion of the joint genotypes *A/*A and *A/*B carrying the ADA₁*2 allele in healthy newborns shows an intermediate value between blood donors and immunological diseases. The data are compatible with the hypothesis that subjects with the genotypes *A/*A and *A/*B carrying the ADA₁*2 allele are more susceptible to post-natal diseases, thereby resulting in the exclusion of them from the possibility to be accepted as blood donors. Based on the difference observed between newborns and blood donors, we have calculated that the proportion of subjects unable to be accepted as blood donors because ADA₁-ACP₁ interaction is around 3%.

Table 4 shows the proportion of ACP₁*A/ADA₁*2 gametic type in relation to past malarial morbidity in the central area of Sardinia. The proportion of this gametic type is decreasing with increasing malarial morbidity thereby suggesting an increased susceptibility to severe manifestations of malaria in

carriers of this gametic type resulting in a negative selection in areas of high endemia.

The observations reported in tables 2-4 suggest that the joint genotypes *A/*A and *B/*A carrying the ADA₁*2 allele are more susceptible to diseases involving the immune system as compared to other genotypes. The following data point to a positive role of this joint genotype on intrauterine growth and against some classes of diseases of post-reproductive period of life.

Fig. 2 shows the proportion of newborns with birth weight $\geq 90^{\circ}$ centile, in relation to ACP₁ and ADA₁ genotypes in a sample of consecutive newborns (1). The proportion of these infants is higher in *A/*A and *A/*B genotypes carrying the ADA₁*2 allele than in other joint genotypes suggesting a favorable effect of these joint genotypes on intrauterine growth.

Table 5 shows the effect of the joint ACP₁-ADA₁ genotype on glycemic level in subjects with type 2 diabetes. The proportion of *A/*A and *A/*B subjects carrying the ADA₁*2 allele is lower in subjects with very high glycemic level than in subjects with a moderate increase of glycemic level. This finding suggests a protective effect of the joint *A/*A and *A/*B genotypes carrying the ADA₁*2 allele against extreme blood glucose levels. Since low ACP₁ activity (*A/*A and *A/*B activity) increases glucose tolerance and this is very marked in subjects carrying the ADA₁*2 allele the association could be related to the lowering of ACP₁ activity in the presence of ADA₁*2 allele, thus explain the highly significant interaction between ADA₁, ACP₁ and glycemia. A cooperative effect appears unlikely considering that a high adenosine concentration due to low ADA activity in ADA₁*2 carriers would bring about a high activity of adenosine receptors thereby resulting in a decreased glucose tolerance (see table 1). The increased glucose tolerance in low ACP₁ activity genotypes could be due to increased glucose metabolism connected with lower dephosphorylation of insulin receptor and B3P but also to decreased dephosphorylation of FMN that may increase the metabolic output due to increased activity of flavoenzymes in Krebs cycle.

Table 6 shows the relationship between the joint ACP₁/ ADA₁ genotypes and Left Ventricular Ejection Fraction (LVEF) in subjects with cardiovascular diseases. Both ACP₁ and ADA₁ activity have been found associated with cardiovascular diseases [27,28]: low ACP₁ activity and low ADA₁ genotypes have been found to exert a protective effect. The table shows that LVEF is significantly higher in the joint genotypes ACP₁ *A/*A and *A/*B carrying the ADA₁*2 allele than in other joint genotypes. This finding may represent an

example of cooperative interaction between the two systems.

Discussion

The finding described by Lucarini et al (2) indicates that in carriers of low activity ACP₁ *A/*A and *A/*B genotypes the presence of ADA₁*2 allele leads to a further decrease of enzymatic activity and this could be responsible for the associations observed at clinical level. However considering the effects of ACP₁ and ADA₁ on the immune response and on glucose metabolism a cooperative action is also possible (Table 1,[3,4]) . A cooperative action on immune reaction is supported by the well known relationship between ADA₁ and immune diseases and by the relationship between ACP₁ and T-cell activation. A cooperative action on metabolic functions is also possible and is suggested by the role of ACP₁ and adenosine receptors activity on glucose metabolism. As a protein phosphotyrosine phosphatase, ACP₁ may have an important role in modulation of glycolytic rate through the control of insulin receptor activity and of band 3 protein phosphorylation status. Catalysing the conversion of FMN in riboflavin, the enzyme may have an important role in regulating flavo-enzyme activity and energy metabolism. Concerning ADA₁, recent studies have shown that adenosine counteracts insulin action in the liver by activating A2B receptors. Concerning ACP₁ possible effects on energy metabolism is also suggested by the action on ACP₁ on FMN.

The observations in tables 2 and 5 are compatible with the hypothesis of a further decreased activity of ACP₁ *A/*A and *B/*A genotypes in carriers of ADA₁*2 allele resulting in negative effects on ZAP70 and in turn on lymphocyte activity thus explaining the increased susceptibility to immunological diseases and malaria. Comparing the proportion of the joint genotype *A/*A and *B/*A carrying the ADA₁*2 allele observed in blood donors with that observed in newborns (fig 1) we have calculated that the effects of these joint genotypes on the health of individuals results in the exclusion of about 3% of the population from the category of blood donors.

A positive role of these joint genotypes has been observed in intrauterine growth, type 2 diabetes and subjects with cardiovascular diseases. The effects on intrauterine growth and on glycemic levels in T2D may be explained by the decrease of on ACP₁ *A/*A and *B/*A activity in carriers of ADA₁*2 allele resulting in an increase of insulin action, glucose utilization and metabolic activity. Since it has been observed that both low activity ACP₁ genotypes and low ADA₁ activity exert a protective effect against coronary artery diseases [26,27], a cooperative effect

of these genotypes on cardiac function appears likely.

Some samples are small and the results need to be confirmed before drawing definitive conclusions.

A statistically significant association between ACP₁ and ADA₁ has been found in Caucasians living in Australia [28] and in a Brazilian population [29]. On the other hand, no significant association has been observed in other populations [30], suggesting some effects of genetic background and/or environmental conditions.

The frequency of ACP₁*A allele in human populations increases with decreasing distance from the North Pole [31]. Observations by our group suggest that the relative intrauterine survival rate of *A/*A genotype carrying the ADA₁*2 allele is greater at high than at low altitude [32]. Since the activity of ACP₁*A/*A is lower in the presence of ADA₁*2 than in its absence, the pattern observed represents a further argument in favor of the hypothesis that low ACP₁ activity has a selective advantage at low environmental temperatures.

Interactions between ACP₁ and ADA₁ with diverse intensity and direction of selective effects during intrauterine and reproductive stages of life would result in a statistical association of diverse strength and direction between the two genetic systems in later stages of life among human populations. Such effects persisting for generations may result in statistical correlations between gene frequencies at corresponding loci. In this context, the observation of Farabegoli and Barbujani [33] that the axis of the gradient for ACP₁ alleles, parallels the gradient observed for ADA locus, is of interest.

The joint system ACP₁-ADA₁ seems to have an important role in human health and development. The ACP₁ (*A/*A and *B/*A) genotypes carrying the ADA₁*2 allele may be advantaged during intrauterine life through a positive effect on fetal growth. During pre-reproductive and early reproductive stages of extra uterine life, individuals carrying these joint genotypes may be disadvantaged since they could be more susceptible to diseases involving the immune system. In the post-reproductive period of life these genotypes may be relatively protected against common diseases occurring during the last period of human life.

The observation on Type 2 diabetes is in agreement with experimental data showing that the reduction of ACP₁ activity increases glucose tolerance [34], thereby pointing to possible beneficial effects of a pharmacologic reduction of ACP₁ activity in diabetic patients.

References

- [1] Gloria-Bottini F, Lucarelli P, Amante A, et al. Interaction at clinical level between erythrocyte acid phosphatase and adenosine deaminase genetic polymorphisms. *Hum Genet* 1989; 82:213-5.
- [2] Lucarini N, Borgiani P, Ballarini P, et al. Erythrocyte acid phosphatase (ACP1) activity. In vitro modulation by adenosine and inosine and effects of adenosine deaminase (ADA) polymorphism. *Hum Genet* 1989; 81:185-7.
- [3] Bottini N, Gloria-Bottini F, Borgiani P, et al. Type 2 diabetes and the genetics of signal transduction: a study of interaction between adenosine deaminase and acid phosphatase locus 1 polymorphisms. *Metabolism* 2004; 53: 995-1001.
- [4] Nicotra M, Bottini N, La Torre M, et al. Repeated spontaneous abortion. Cooperative effects of ADA and ACP1 genetic polymorphisms. *Am J Reprod Immunol.* (2007); 58:1-10.
- [5] Ammendola M, Pietropolli A, Saccucci P, et al. Acid phosphatase locus 1 genetic polymorphism, endometriosis, and allergy. *Fertil Steril* 2008; 90: 1203-5.
- [6] Saccucci P, Manca Bitti ML, Bottini N, et al. Type 1 diabetes: evidence of interaction between ACP1 and ADA1 gene polymorphisms. *Med Sci Monit* 2009; 15:511-517.
- [7] Gloria-Bottini F, Saccucci P, Magrini, A et al. Is there a role of ACP1-ADA1 genetic complex in immune reaction? Association with T1D and with past malarial morbidity. *Am J Med Sci* 2010; 340:268-70.
- [8] Bottini N, Bottini E, Gloria-Bottini F, et al. Low-molecular-weight protein tyrosine phosphatase and human disease: in search of biochemical mechanisms. *Arch Immunol Ther Exp (Warsz)* 2002; 50:95-104.
- [9] Bottini N, Stefanini L, Williams S, et al. Activation of ZAP-70 through specific dephosphorylation at the inhibitory Tyr-292 by the low molecular weight phosphotyrosine phosphatase (LMPTP). *J Biol Chem* 2002; 277:24220-4.
- [10] Chiarugi P, Cirri P, Marra F, et al. LMW-PTP is a negative regulator of insulin-mediated mitotic and metabolic signalling. *Biochem Biophys Res Commun* 1997; 238:676-82.
- [11] Shekels LL, Smith AJ, Van Ethen RL, et al. Identification of the adipocyte acid phosphatase as a PAO-sensitive tyrosyl phosphatase. *Protein Sci* 1992; 1:710-21.
- [12] Londos C, Honnor RC, Dhillon GS. cAMP-dependent protein kinase and lipolysis in rat adipocytes. III. Multiple modes of insulin regulation of lipolysis and regulation of insulin responses by adenylate cyclase regulators. *J Biol Chem* 1985; 260:15139-45.
- [13] Ohisalo JJ, Strandberg H, Kostiaainen E, et al. Stimulation of lipoprotein lipase activity of rat adipose tissue and post-heparin plasma by N6-(phenylisopropyl)adenosine. *FEBS Lett* 1981; 132:121-3.
- [14] Vannucci SJ, Nishimura H, Satoh S, et al. Cell surface accessibility of GLUT4 glucose transporters in insulin-stimulated rat adipose cells. Modulation by isoprenaline and adenosine. *Biochem J* 1992; 288:325-30.
- [15] Challis RA, Budohoski L, McManus B, et al. Effects of an adenosine-receptor antagonist on insulin-resistance in soleus muscle from obese Zucker rats. *Biochem J* 1984; 221: 915-7.
- [16] Challis RA, Richards SJ, Budohoski L. Characterization of adenosine receptor modulating insulin action in skeletal muscle. *Eur. J. Pharmacol* 1992;226:121-128.
- [17] Franco R, Casadó V, Ciruela F, et al. Cell surface adenosine deaminase: much more than an ectoenzyme. *Prog Neurobiol* 1997; 52: 283-94.
- [18] Franco R, Aran JM, Colomer D, et al. Association of adenosine deaminase with erythrocyte and platelet plasma membrane: an immunological study using light and electron microscopy. *J Histochem Cytochem* 1990; 38:653-8.
- [19] Dong RP, Kameoka J, Hegen M, et al. Characterization of adenosine deaminase binding to human CD26 on T cells and its biologic role in immune response. *J Immunol* 1996; 156:1349-55.
- [20] Ciruela F, Saura C, Canela EI, et al. Adenosine deaminase affects ligand-induced signalling by interacting with cell surface adenosine receptors. *FEBS Lett* 1996; 380:219-23.
- [21] Xu B, Berkich DA, Crist GH, et al. A1 adenosine receptor antagonism improves glucose tolerance in Zucker rats. *Am J Physiol* 1998; 274:271-9.
- [22] Richardson PJ. Asthma. Blocking adenosine with antisense. *Nature* 1997; 385:684-5.
- [23] Yasuda N, Inoue T, Horizoe T, et al. Functional characterization of the adenosine receptor contributing to glycogenolysis and gluconeogenesis in rat hepatocytes. *Eur J Pharmacol* 2003; 459:159-66.
- [24] Kameoka J, Tanaka T, Nojima Y, et al. Direct association of adenosine deaminase with a T cell activation antigen, CD26. *Science* 1993; 261: 466-9.
- [25] Wurzinger KK, Novotny JE, Mohrenweiser HW. Studies of the purine analog associated modulation of human erythrocyte acid phosphatase activity. *Mol Cell Biochem* 1985; 66:127-136.
- [26] Safranow K, Rzeuski R, Binczak-Kuleta A, et al. ADA*2 allele of the adenosine deaminase gene may protect against coronary artery disease. *Cardiology* 2007; 108:275-81.
- [27] Banci M, Saccucci P, D'Annibale F, et al. ACP1 genetic polymorphism and coronary artery disease: an association study. *Cardiology* 2009; 113:1236-42.
- [28] Brackenridge C. A ternary-associating system of genes. *Hum Hered* 1997;27:114-117
- [29] Engrácia V, Mestriner MA, Cabello PH, et al. Association between the acid phosphatase I and adenosine deaminase systems in a Brazilian sample. *Hum Hered* 1991; 141 :147-50.
- [30] Hernández JL, Elston RC, Ward LJ. Gametic equilibrium between 24 polymorphic markers. *Hum Genet* 1990; 85:343-8.
- [31] Ananthakrishnan R, Walter H. Some notes on the geographical distribution of the human red cell acid phosphatase phenotypes. *Humangenetik* 1972;15:177-181
- [32] Gloria-Bottini F, Lucarelli P, Lucarini N, et al. Adenosine deaminase-acid phosphatase association and the environment: A study in a continental Italian population. *Am J Hum Biol* 2000; 12:214-220.
- [33] Farabegoli A, Barbujani G. Diversity of some gene frequencies in European and Asian populations. VI. Geographic patterns of PGM and ACP. *Hum Hered* 1990; 40:313-21.
- [34] Pandey SK, Yu XX, Watts LM, et al. Reduction of low molecular weight5 protein-tyrosine phosphatase expression improves hyperglycemia and insulin sensitivity in obese mice. *J Biol Chem* 2007; 282:14291-14299.

Table1. Mechanism by which ADA₁ and ACP₁ activity may influence immune response and glucose metabolism

		Effect on immune response	Effect on glucose metabolism
Low concentration of adenosine due to high ADA activity(ADA ₁ *1/*1)	Low activity of adenosine receptors	Decreased T cell activity	Increased glucose tolerance
High concentration of adenosine due to low adenosine deaminase activity (carriers of ADA ₁ *2 allele)	High activity of adenosine receptors	Increased T cell activity	Decreased glucose tolerance
Low dephosphorylating activity on the negative regulatory Tyr 292 of ZAP70 and on tyrosine kinase receptors (ACP ₁ *A*A and *A/*B)	Low ZAP-70 activity and high activity of tyrosine kinase receptors	Decreased T cell activity	Increased glucose tolerance
High dephosphorylating activity on the negative regulatory Tyr292 of ZAP70 and on tyrosine kinase receptors (other ACP ₁ genotypes)	High ZAP-70 activity and low activity of tyrosine kinase receptors	Increased T cell activity	Decreased glucose tolerance

Table 2. Proportion of low activity ACP1 genotypes *A/*A and *A/*B in ADA1*1/*1 subjects and in carriers of ADA*2 allele. Comparison between blood donors and subjects with immunological diseases.

		%proportion of *A/*A and *A/*B	Total n°	O.R.
IMMUNOLOGICAL DISEASES				
Types 1 diabetes Children	ADA ₁ *1/*1	47.4%	232	2.21
	Carriers of ADA ₁ *2	66.6%	54	
Types 1 diabetes Adults	ADA ₁ *1/*1	37.1%	89	1.70
	Carriers of ADA ₁ *2	50.0%	18	
Women with repeated spontaneous abortion	ADA ₁ *1/*1	49.0%	153	2.50
	Carriers of ADA ₁ *2	70.6%	17	
Allergic women with endometriosis	ADA ₁ *1/*1	36.7%	30	2.90
	Carriers of ADA ₁ *2	62.5%	8	
Crohn's disease	ADA ₁ *1/*1	28.3%	53	1.77
	Carriers of ADA ₁ *2	41.2%	17	
All diseases	ADA ₁ *1/*1	43.5%	557	1.97
	Carriers of ADA ₁ *2	60.5%	114-	

BLOOD DONORS				
Sample 1°	ADA ₁ *1/*1	49.1%	171	0.26
	Carriers of ADA ₁ *2	20.0%	30	
Sample 2°	ADA ₁ *1/*1	43.3%	67	0.38
	Carriers of ADA ₁ *2	22.7%	22	
All blood donors	ADA ₁ *1/*1	47.5%	238	0.30 95% C.I. 0.14 – 0.63
	Carriers of ADA ₁ *2	21.1%	52	
Statistical analysis : Three way contingency table analysis by a log linear model X=ACP ₁ y=ADA ₁ z=Sample				
		G	df	p
IMMUNOLOGICAL DISEASES				
	xyz interaction	0.526	4	0.970
	xy independence	13.249	5	0.025
	xz independence	15.635	8	0.070
	yz independence	12.850	8	0.120
BLOOD DONORS				
	xyz interaction	0.285	1	0.650
	xy independence	12.546	2	0.002
	xz independence	0.716	2	0.700
	yz independence	3.471	2	0.180
ALL IMMUNOLOGICAL DISEASES VS ALL BLOOD DONORS				
	xyz interaction	22.948	1	<0.0002
	xy independence	23.506	2	<0.0001
	xz independence	24.206	2	<0.0001
	yz independence	23.094	2	<0.0001

Table 3. Proportion of ACP₁ *A/*A and *A/*B genotypes carrying the ADA₁*2 allele. Comparison between blood donors and subjects with immunological diseases

	%Proportion of *A/*A and *A/*B subjects carrying ADA ₁ *2 allele	Total n°
IMMUNOLOGICAL DISEASES		
Types 1 diabetes Children	12.6%	286
Types 1 diabetes Adults	8.4%	107
Women with repeated spontaneous abortion	7.1	170
Women with allergic endometriosis	13.1%	38
Crohn’s disease	10.0%	70
All diseases	10.3%	671
<i>Chi square test of independence</i>		
	χ^2 4.314	<i>df</i> 4
		<i>p</i> 0.365
BLOOD DONORS		
Sample 1°	3.0%	201
Sample 2°	5.6%	89
All blood donors	3.8%	290
<i>Chi square test of independence</i>		
	χ^2 0.561	<i>df</i> 1
		<i>p</i> 0.454
<i>Comparison all immunological diseases vs all blood donors</i>		
	10.341	1
		0.0013
<i>O.R. 2.91</i>	<i>95% C.I. 1.46-5.91</i>	

Table 4 . Proportion of ACP₁ *A/ADA₁*2 gametic type in relation to past malarial morbidity in the central area of Sardinia (from Ref. 7 modified).

	malarial morbidity %	Altitude –meters above sea level	Proportion of *A/ADA ₁ *2 gametic type	Total n° of gametic types
Low altitude	82%	10-45	0.4%	786
Medium altitude	29%	185-648	1.1%	1262
High altitude	17%	796-1000	1.8%	720
<i>Chi square test of independence</i>			χ^2 7.188	<i>df</i> 2
				<i>p</i> 0.027

Table 5. The effect of ACP₁ and ADA₁ on glycemic level in T2D.

Glycemia	ACP ₁ genotype	Proportion of ADA ₁ *2 carriers	Total n°
≤160	*A/*A and *A/*B	19.8%	96
	Other genotypes	15.8%	120
>160	*A/*A and *A/*B	4.8%	21
	Other genotypes	31.6%	36
<i>Three way contingency table analysis</i> x=ADA ₁ ; y=ACP ₁ ; z=Glycemia			
xyz interaction		p=0.001	

Table 6. Left Ventricular Ejection Fraction in subjects with cardiovascular diseases

ACP1 genotype	Ejection Fraction		
	Mean	S.E.	Total n°
*A/*A and *A/*B genotypes carrying ADA ₁ *2 allele	57.29	1.34	17
Other joint genotypes	51.93	0.52	322
Student-t for difference between means			
	t	p	
	3.747	0.001	

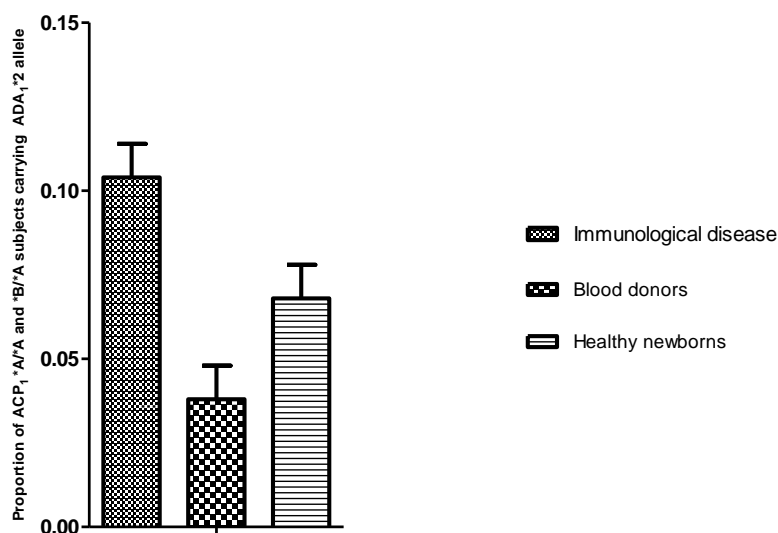


Fig.1. Proportion of ACP₁ *A/*A and *B/*A genotypes carrying the ADA₁*2 allele. Comparison among patients with immunological diseases, healthy adults and healthy newborns from the same population: p=0.002.

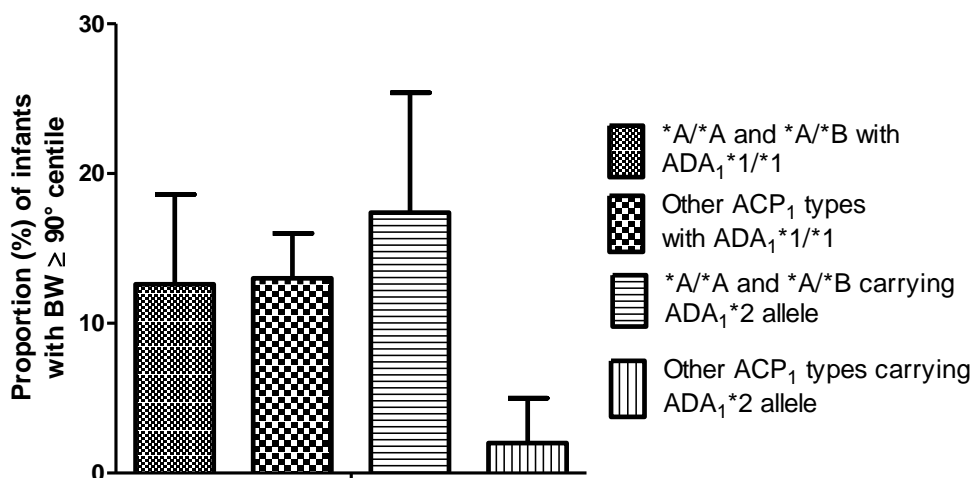


Fig 2. Proportion (%) of newborns with BW ≥ 90th centile in relation to ACP₁ and ADA₁ genotypes. Interaction between ACP₁, ADA₁ and BW (p<0.02).