The Effects of Aerobic Exercise and Melatonin on CA Activity in Rats

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Abstract: The purpose of this study was to determine the effects of Aerobic exercise and Melatonin on carbonic anhydrase activity in rats. In this study, 24 healthy Spraque Dawley male rats, weighing 250–300 g, 4–6 months of age were used. The rats were divided into four equal groups. C; (Control Group), M; (Melatonin Group), E; (Exercise Group), ME; (Melatonin-Exercise Group). The exercise groups (n:12) were run for endurance training at 23°C. The exercise was carried out 6 days a week for 4 weeks. The intensity of endurance training was gradually increased during four weeks. A standard mild electric shock deterrent was used intermittently when necessary to coerce the rats to run at the end of the fourth week, rats were killed by decapitation on each training day. According to the result of this study, the level of carbonic anhydrase decreases in control, melatonin and exercise groups and melatonin decreases significantly in exercise group. As a result, it can be said that melatonin and exercise reduces the level of CA in mechanisms related to O2 consumption and CO2 production in organism by having an organizer role.

Keywords: Melatonin, Carbonic anhydrase, Aerobic Exercise, Rat

INTRODUCTION

Melatonin, the pineal gland hormone, is secreted in a characteristic circadian rhythm, with its main production occurring during the dark phase. This endogenous rhythm is driven by the suprachiasmatic nuclei, which regulate the activity of N-acetyltransferase (NAT), the rate-limiting enzyme in melatonin biosynthesis. In addition, NAT activity is immediately suppressed by light affecting the retina through a monosynaptic pathway [1]. An increase in Melatonin after strenuous physical exercise has been reported in man, but the results were inconsistent. Recently, an increase in Melatonin was found in rats that had been exposed to hypobaric hypoxia [2]. Moreover, it is considered as a powerful free radical scavenger and likely to be a general promoter of antioxidative mechanisms and apotential antioxidant in vitro and in vivo [1].

The original and primary function of melatonin in organisms is to serve as an antioxidant to detoxify the free radicals generated during the process of aerobic metabolism with the other functions of melatonin [3-4]. In addition, clinical trials in humans have reported that melatonin effectively reduces the severity of diseases and disorders in which their etiologies are associated with free radical damage or oxidative stress [5-6]. While the results of these experimental and clinical studies clearly document the ability of melatonin to reduce oxidative damage resulting from a variety of oxidative insults in widely different organs, the possibility exists that the protective actions of the indole are related exclusively to its radical scavenging activities as well as to its ability to work in conjunction with other antioxidants via the network described above [7].

Melatonin is known to influence a variety of biological processes including circadian rhythms, neuroendocrine, cardiovascular and immune functions as well as thermoregulation [8-9].

In rats, melatonin through its antioxidant properties, protected the animals from adriamycin-induced nephropathy and cardiomyopathy [10]. Melatonin, because of its antioxidant properties and lipophilic and hydrophilic nature, may be applicable in the treatment of disorders in which oxidative stress is involved, including exercise-induced oxidative stress. The aim of our study is to examine the effect of melatonin in exercise-induced oxidative stress in healthy subjects. Exercise is known to exert numerous physiological changes in vital organ system of the body. Among those changes, the most important is the enhanced respiration and utilization of oxygen in the body. Increased oxygen influx
During exhaustive exercise may be potentially harmful to the body. During the last 10 years, much evidence has accumulated implicating enormous generation of reactive oxygen species and other free radicals especially during exercise in the muscles and heart [11]. Although the exact mechanism for the exercise-induced cell and tissue damage is still elusive, there is an increasing evidence that the enhanced oxidative metabolism associated with the exercise can increase the whole body oxygen consumption 10 to 20 fold [12]. The antioxidant enzyme activities are prone to alteration due to changes in oxygen consumption (oxidative stress). Oxidative stress can be described as a disturbance in the antioxidant system which is not able to adequately scavenge free radicals/reactive oxygen species and arrest lipid peroxidation [11].

Carbonic anhydrase (CA, carbonate hydrolyase, EC 4.2.1.1) is a metalloenzyme. CAs are an important class of enzymes used to regulate CO₂ levels in living organisms by catalysing the reversible hydration of CO₂ to HCO₃⁻ and H⁺ [9-13].

Carbonic anhydrase (CA) isozymes play key roles in diverse processes, such as physiological pH control and gas balance, calcification, and photosynthesis [14]. However, Carbonic anhydrase (CA; Carbonate hydrolyase, EC 4.2.1.1) is a family of metalloenzymes that catalyze the rapid conversion of carbon dioxide to bicarbonate and protons and involved in the biomineralization process [15]. In the biomineralization process, the mineral structures involved are mainly calcium carbonate and calcium phosphate crystals, in invertebrates and vertebrates, respectively. This enzyme is a multifunctional enzyme that catalyzes the hydration/dehydration of carbon dioxide. The molecular characteristics of the CA across the plant and animal breeds are similar. It catalyzes the reversible hydration of CO₂ to HCO₃⁻ and H⁺. In the red blood cell, this enzyme is necessary to facilitate the transport of carbon dioxide out of the body [14-16]. Carbonic anhydrase greatly increases the rate of the reaction, with typical catalytic rates of the different forms of this enzyme ranging between 104 and 106 reactions per second. The active site of most carbonic anhydrases contains a zinc ion. Many natural and synthetic substances can affect the living metabolism by altering enzyme activities and affecting metabolic pathways at low concentrations [17]. CA plays an important role in water and ion transport and pH regulation in kidney, eye, central nervous system (CNS), inner ear and other systems [15].

So far, many studies have been carried out on the effects of exercise and Melatonin on variables such as growth hormone, etc., but no study has been done on the effects of aerobic exercise and melatonin on carbonic anhydrase activity. Therefore, in the present study we examined the effects of aerobic exercise and melatonin which had previously been shown to protect cellular components from free radical damage on Carbonic anhydrase activity in rats.

MATERIAL AND METHODS

Chemicals

The used melatonin is provided from sigma factory in the form of 1g prepare dust. After the dissolution of melatonin in 0.5% Ethanol rate and by addition of 0.9% Sodium Chloride, all of the appropriated melatonin groups, were prepared in 50 ml solution form. 1-1.5 hour before beginning of exercise and darkness, daily doses (5 mg/kg) were given intraperitoneally during 4 weeks.

Animals and groups

In this study, 24 healthy Spraque Dawley male rats, weighing 250-300 g, 4-6 months of age, provided from Firat University Experimental Animal Research Center (FUDAM). The study was carried out in Ataturk University Research Center of Experimental Animals and the study was approved by the Ethical Committee of the Ataturk University (AUAHYEKE, Ethical Committee Report No: 2008-51). All surgical procedures and protocols used here were in accordance with Guidelines for Ethical Care of Ataturk University Research Center of Experimenta Animals.

The rats were kept under special conditions and were sheltered in cages, each with 6 rats, at the room temperature (25°C), supplying with food (Bayramoğlu Yem Sanayi, Erzurum, Turkey) and water for 12–hour day and night cycles. The rats were divided into four equal groups.

C; (Control Group), The sedentary group on which no application was employed.
MC; (Melatonin-Control Group), The ones that were given melatonin, but none were made exhaustive exercise.
E; (Exercise Group), The ones that were made exhaustive exercise.
ME, (Melatonin-Exercise Group), The ones that were given melatonin and were made exhaustive exercise.

Exercise Protocol

The same exercise programs were applied to exercise and melatonin exercise groups in test groups. During the test, digital thermometer (GEMO, micro software and PID thermo controlled device) was used.

Adaptation Training: For the rats to have adaptation they were made to have exercise on treadmill for 10-15 min, at 20 m/min during per week (MAYTME 9805 treadmill exerciser, Commat Ltd., Ankara, Turkey). Adaptation training was made at 23°C laboratory temperature.

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Loading Training: The exercise groups (n:12) were exposed to endurance training in laboratories at 23оС. The exercise was done 6 days a week during 4 weeks. Endurance training was gradually increased by increasing the training of speed and time during four weeks. A standard mild electric shock deterrent was used intermittently when necessary to coerce the rats to run. At the end of the fourth week, rats were killed by decapitation on each training day.

The physical training program:
First week: Time: 30 min; Speed; (23 m/min.)
Second week: Time: 40 min; Speed; (23 m/min.)
Third week: Time: 50 min; Speed; (25 m/min.), (2 % slope)
Fourth week: Time: 60 min; Speed; (25 m/min.), (2 % slope)

Drawing of blood and preparation of haemolysate
Venous blood was drawn from the vena cava inferior into a sterile plastic syringe (10 mL) using a sterile needle. Half of the drawn blood (3 mL) was added to a plastic test tube containing 50 µL of EDTA (1:100) to be used for the carbonic anhydrase enzyme activity assay. Erythrocytes were isolated from fresh rat blood after exhaustive exercise and hypothermic stress. Immediately, the fresh blood was centrifuged at low-speed centrifugation (1500 rpm) for 15 min (HERMLE Z 323 K) by removal of plasma anduffy coat. The erythrocyte pellet was washed three times with cold 0.16 M KCl and the supernatant discarded. One volume of erythrocyte pellet was suspended in five volumes of ice water to give an erythrocyte haemolysate. CA activity was determined colorimetrically as described above [18-19-20].

Protein determination
Quantitative protein determination was achieved by absorbance measurements at 595 nm according to Bradford’s method (1976), with bovine serum albumin as standard described previously [21-22].

Hemoglobin estimation
The hemoglobin (Hb) concentration in hemolysate was determined by the cyanmethaemoglobin method. All studies were performed at +4оС [21-23].

Carbonic anhydrase enzyme activity determination
Carbonic anhydrase activity was assayed by following the hydration of CO₂ at room temperature according to the method described by Wilbur and Anderson (1976). CO₂-hydratase activity as an enzyme unit (EU) was calculated by using the equation (t₀−tₖ/tₖ) where t₀ and tₖ are the times for pH change of the nonenzymatic and the enzymatic reactions, respectively [14].

Statistical analysis
The experimental results were performed in triplicate. The data were recorded as mean ± standard deviation and analyzed by SPSS (version 11.5 for Windows 2000, SPSS Inc.). For determining the mean of two groups different from each other, the Mean-Whitney U test which is a non-parametric test (P<0.05) regarded as significant, and P<0.001, very significant is used.

3. RESULTS
CA Levels:

![Figure 1](http://www.ijSciences.com)  
Figure 1. In the graph in which CA levels of Control, Melatonin, Exercise And Melatonin-Exercise Groups are compared, the level of carbonic anhydrase in blood in melatonin-exercise groups is significantly low compared to other groups (*;P<0.05), and the highest level of CA was determined in control group which no application was applied. Data are shown as the mean±S.D. for each group.
4. DISCUSSION
Melatonin exists in all body fluids after its release from the pineal gland. It is often utilized, as a sleep-inducing agent being an important hormone. The pineal hormone has influence on a variety of biological processes including circadian rhythms, neuroendocrine, cardiovascular and immune functions, as well as for thermoregulation [14]. Additionally, melatonin functions in protecting cell components such as nuclear DNA, membrane lipids and cytosolic proteins from free radical damage [24]. Many chemicals when administered at relatively low doses affect metabolism by altering normal enzyme activity, particularly through inhibition of a specific enzyme. The effects can be dramatic and systemic [25-26]. Therefore, melatonin has an important role by altering the activities of enzymes used to improve the total antioxidative defence capacity of the organism [27]. Its impact on carbonic anhydrase (CA) activity has not previously been reported.

Carbonic anhydrase is a very important enzyme in the body and all the CA isozymes are deeply involved in a great number of secretory activities including fluid movements [9-20]. The physiological function of the CA isozymes is important in facilitating the interconversion of CO2 and HCO3; also, they play key roles in physiological pH control in most tissue [28]. Some studies have examined the effects of Melatonin, for example, Beydemir and Gulcin, studied reviews the effects of Melatonin on Carbonic Anhydrase from Human Erythrocytes In Vitro and from Rat Erythrocytes In Vivo. They observed that CA activity in the rat erythrocytes was decreased by the melatonin after 1 and 3 hours. However, CA activity was restored to its normal level after 6 h. probably due to metabolism of the melatonin. The findings indicate that melatonin may be pharmacologically useful in some diseases. HCA-I and HCA-II were purified from human erythrocytes. CA is widely distributed in most tissues and has a very important role in some diseases such as glaucoma, where CA inhibitors are generally used for treatment since they reduce intraocular pressure. Our study revealed that there was a good correlation between the in vivo and in vitro inhibitory effects of melatonin on HCA-I and HCA-II activities [14].

In another study Kumar et al. studied the effect of oral melatonin on exercise-induced oxidant stress in healthy subjects. The result showed Lipid peroxidation products measured as malondialdehyde – equivalents (MDA-eq) were significantly increased whereas superoxide dismutase and glutathione peroxidase were decreased in subjects after the exercise. There was no change in the total antioxidant activity in plasma or catalase activity in RBC before and after the exercise. The basal levels of lipid peroxidation products were significantly decreased in subjects treated with melatonin as compared with the study without melatonin. The total antioxidant activity was significantly increased in plasma of subjects treated with melatonin. Exercise-induced reduction in superoxide dismutase and glutathione peroxidase was prevented following melatonin treatment [11]. Olcay and colleagues in their study that concluded that Melatonin hormone inhibited the CA of erythrocyte in vivo. Therefore, it may demolish physiological events, i.e. preservation red cell intracellular, pH, ventilatory control and red cell fragility. For this reason, dosage, duration and methods of administration of melatonin should be further evaluated and excessive applications should be avoided [29].

However, no studies have examined the effects of aerobic exercise and melatonin on carbonic anhydrase activity in rats. Therefore, in this study, the effects of Aerobic exercise and Melatonin on Carbonic anhydrase activity in rats were determined. As a result, the level of carbonic anhydrase decreases significantly in melatonin-exercise group compared to control, melatonin and exercise and melatonin groups. We can say that melatonin and aerobic exercise raise the level of CA, which has a protective part in O2 consumption and CO2 production in organisms and in transferring of CO2 out of tissues and balancing the levels of acid and base in blood and other tissues.

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