

## Effect of ATF5 in Mice Transfected with Human Cytomegalovirus IE2 Gene

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**Abstract:** To investigate the qualitative and quantitative detection of ATF5 activated transcription factors in tumor tissue by using the method of transfecting IE2 mouse tumor to explore the effect of HCMV immediate early protein IE86 on ATF5 function. Transfected mice were treated with GL261 mouse glioma cells. The expression of transcription factor ATF5 in the tumor tissues of tumor-bearing mice was detected by PCR and Western-blotting methods. The experimental results showed that the expression of ATF5 was higher in IE2 transgenic mice than in normal mice. ATF5 is highly expressed in IE2 transgenic mice. ATF5 is a transcription factor closely related to apoptosis and plays an important role in apoptosis, differentiation and development.

**Keywords:** Human Cytomegalovirus, IE86, Mouse Glioma Cell, ATF5

### Introduction

HCMV belongs to the herpes virus family and causes widespread and persistent infections in the human population. The brain is a preferred site for HCMV infection, leading to dysfunction such as mental disorders or epilepsy [1-2]. HCMV regulates the malignant phenotype of the tumor by affecting cell survival, cell cycle and invasion potential. The IE2 gene mouse was established by DNA pronuclear microinjection method[3]. The target gene transferred is only integrated into one of the chromosomes of the diploid animal, so it is a hemizygous, and its offspring only have a part of the individual. There are integrated genes, some have no target genes, so transgenic mice require a fairly long screening and purification process to obtain homozygous transgenic mice. The first viral gene after HCMV infection is an immediate early gene, and the most abundantly expressed product in the gene is called immediate early 1 and 2 proteins [4] (IE72 and IE86). The IE86 protein is a potent transactivator that interacts with factors of the underlying transcriptional machinery. The IE86 protein involved in the transactivation of viral and cellular promoters is unique in viral regulatory proteins because it regulates both viral and cellular promoters both negatively and positively [5]. There is increasing evidence that ATF5 is a factor closely related to apoptosis. According to experiments, the interference inhibiting the function of ATF5 in the murine glioma model causes specific apoptosis of glioma cells, but does not have such apoptotic effect on normal cells surrounding glioma cells[6]. High expression of ATF5 has also been found in many tumor tissues, for example, interference in the

inhibition of ATF5 expression in breast cancer cells can also cause apoptosis in cells [7]. ATF5 is a transcription factor closely related to apoptosis and plays an important role in physiological processes such as apoptosis, differentiation and development.

### MATERIALS AND METHODS

#### Animals and Cell culture

Thirty C57BL/6 mice, female, body weight 18-20g, 6-8 weeks old; GL261 mouse glioma cells were cultured in DMEM medium containing 10% fetal calf serum containing penicillin and streptomycin, cultured in 37 °C, 5% CO<sub>2</sub> incubator, subcultured every three days for logarithmic growth phase. The cells were subjected to the following experiments. When the cells were grown to 90% confluence in the culture flask, the cells were digested with trypsin containing EDTA, and the cells were digested into 2 ml of 1×10<sup>7</sup>/ml suspension cells; the mice of the three experimental groups were given the right side of the experimental group by syringe. Under the injection, the control group was injected with the same dose of suspension cells at the same position.

#### RNA extraction of tumor tissue and RT-PCR identification of ATF5

RNA was extracted from each tissue of thirty transgenic mice using RNApure tissue Cell Kit (CWBIO, China). And 1 μg RNA was reverse-transcribed using the Transcriptor First Strand cDNAs Synthesis Kit (Roche, Germany) to produce cDNAs according to the manufacturer's instructions. Primers ATF5-F: ACCTCCACCACCAGCAGCAG

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and ATF5-R: AGCCAGCAGGTCCAAGGTATCC were used to amplify the ATF5 gene with 225 bp product. The band intensity was obtained using Image J software.

#### **Protein extraction of tumor tissue and quantitative detection of ATF5 by Western-blot**

Weigh the tumor tissue with an electronic balance and weigh 20-30 mg into a 1.5 ml autoclaved centrifuge tube, grind thoroughly with a sterile grind, add 501  $\mu$ L of lysate (RIPA:PMSF=500:1), and finally on ice. The cells were lysed for 30 min, centrifuged at 13000 r/min for 4 min at 4 ° C, and the supernatant was collected for total protein. The extracted protein was added to 5 $\times$ SDS loading buffer at a ratio of 1:4, boiled for 5 min, and subjected to SDS-PAGE gel electrophoresis. After SDS-PAGE gel electrophoresis, the protein was transferred from the gel to the PVDF membrane with a constant current of 300 mA. On the top, 5% skim milk powder prepared with TBST was blocked for 2 h, primary antibody (1:2000 dilution) was incubated at 4 °C overnight, secondary antibody (1:2000 dilution) was incubated for 2 h, TBST was washed 3 times for 10 min, and ECL was added. The solution (Form A: B solution = 1:1) was stored in the dark for 5 min and then developed to detect the expression of the target protein.

### **RESULTS**

#### **PCR qualitative identification of ATF5**

The 30 standardized mice of the experiment were identified by tail tail, and the identified negative mice and positive mice were grouped for subsequent experiments. The results showed that the expression of ATF5 in 5 pairs of negative mice and positive mice. Significant differences can be observed by electrophoresis strips (Figure 1).

#### **Tumor status after tumor-bearing in mice**

For the selected tumor-bearing mice, the body weight of the mice was no longer increased, and the mice were sacrificed by cervical dislocation and dissected (Figure 2A). The tumor was stripped and a distinct state difference was observed (Figure 2B). The positive tumor-bearing mice had significant volume difference compared with the negative tumor-bearing mice, and there was a significant difference compared with the control group, which was statistically significant ( $P < 0.0001$ ).

#### **Western blot analysis of quantitative detection of ATF5 expression levels**

Protein expression of tumor-bearing mice was followed by Western blotting to detect the expression of ATF5. Western blot was used to detect the quantitative relationship between ATF5 and IE86 protein (Figure 3A). The experimental group was positive tumor-bearing mice ATF5. The mean value of expression, the control group was the mean value of ATF5 expression in the negative tumor-bearing mice

(Figure 3B), and there was a significant difference compared with the control group, which was statistically significant ( $P < 0.05$ ).

### **Discussion**

Human cytomegalovirus (HCMV) belongs to the beta herpesvirus subfamily and is the largest group of human herpesviruses [8, 9], with an infection rate as high as 100%. The HCMV genome is a double-stranded DNA of more than 240 kb in length, and the entire genome contains 250 open reading frames (ORFs). In the process of viral infection, the expression of HCMV gene shows a certain sequence, which can be divided into early stage (IE), early (E) and late (L) genes [10-12]. After the virus penetrates into the cell, the IE gene is activated by the host cytokine and is expressed first, encoding two important regulatory proteins, IE1 (IE72) and IE2 (IE86). Among them, IE2 (IE86) protein is an important transactivator and plays an important role in HCMV infection [13]. In recent years, it has been confirmed that IE2 (IE86) regulates many factors related to the control of cell cycle.

The IE2 gene mouse used in this experiment is a micronucleus injection method of DNA, and the target gene transferred is only integrated into one of the chromosomes of the diploid animal, so it is a hemizygous, its In the offspring, only a part of individuals have integrated genes, and some have no target genes [14]. Therefore, transgenic mice require a relatively long screening and purification process in order to obtain homozygous transgenic mice. Activating transcription factor 5 (ATF5) was originally isolated from human T lymphocytes and plays an important role in tumor development [15]. ATF5 is a member of the ATF/CREB family of basic leucine zipper (bZIP) domains. Due to the leucine zipper motif and the basic region rich in lysine and arginine residues, ATF5 is involved in DNA binding and protein-protein interactions and has anti-apoptotic effects [16]. The study found that the anti-apoptotic function of ATF5 is selective and has no anti-apoptotic function in the process of apoptosis induced by DNA damage agents. ATF5 is a transcription factor closely related to apoptosis and plays an important role in physiological processes such as apoptosis, differentiation and development [17].

The innovation of this study is to establish animal model of glioma cells under the skin of mice with transgenic mice as an entry point, focusing on the relationship between expression and distribution between IE86 and ATF5 at the molecular level. The results showed that activating transcription factor 5 (ATF5) showed a high expression status in the transgenic IE2 mice, and there was a significant quantitative relationship with it [18-20]. The results of this study are important for the treatment of viral infections and the expansion of new ideas for the treatment and prevention of cancer.

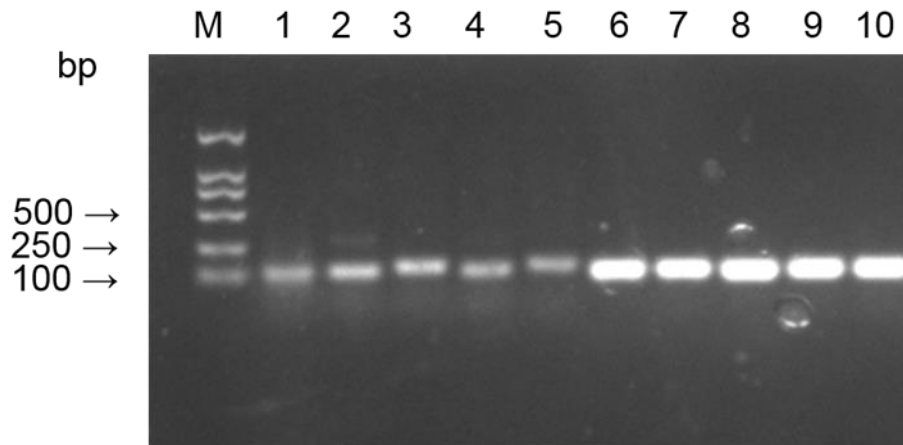


Figure 1 M: DNA marker; 1-5: expression of ATF5 in negative mice, 6-10: Expression of ATF5 in positive mice

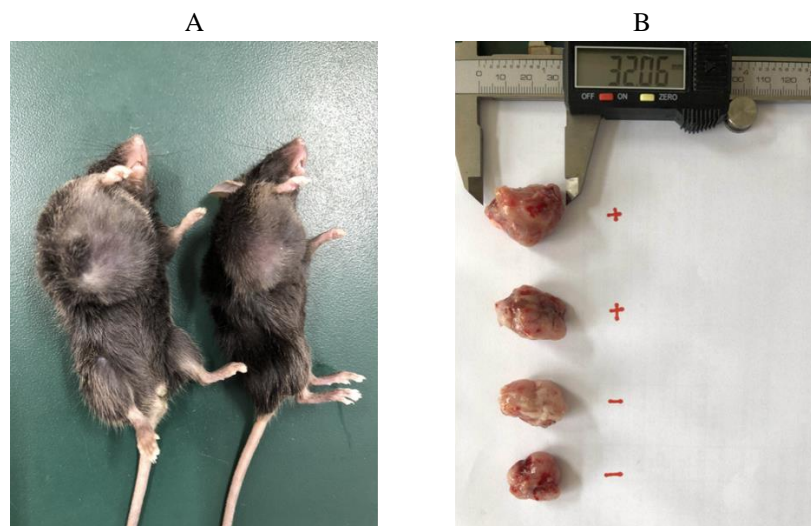


Figure 2 Morphological changes of GL261 murine glioma cells in tumor-bearing mice. In vivo morphological changes (A) and in vitro morphological changes (B).

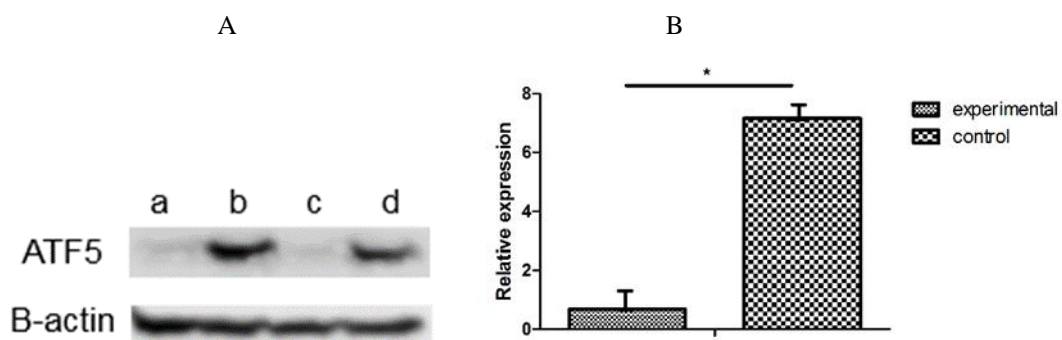


Figure 3 A: The quantitative relationship between ATF5 and IE2 was detected from the protein level by Western blot; B: a, c is the expression level of ATF5 in negativity mice, b, d is the expression level of ATF5 in positive mice, \* P < 0.05

## References

1. Ahn, K., et al., The ER-luminal domain of the HCMV glycoprotein US6 inhibits peptide translocation by TAP. *Immunity*, 1997. 6(5): p. 613.
2. Varnum, S.M., et al., Identification of Proteins in Human Cytomegalovirus (HCMV) Particles: the HCMV Proteome. *Journal of Virology*, 2004. 78(20): p. 10960-6.
3. Zhu, H., Y. Shen, and T. Shen, Human cytomegalovirus IE1 and IE2 proteins block apoptosis. *Journal of Virology*, 1995. 69(12): p. 7960.
4. Arend, K.C., et al., Multiple transcripts encode full-length human cytomegalovirus IE1 and IE2 proteins during lytic infection. *Journal of Virology*, 2016. 90(19): p. JVI.00741-16.
5. Jupp, R., et al., Human cytomegalovirus IE86 protein interacts with promoter-bound TATA-binding protein via a specific region distinct from the autorepression domain. *Journal of Virology*, 1993. 67(12): p. 7539-46.
6. Hansen, M.B., et al., Mouse Atf5: molecular cloning of two novel mRNAs, genomic organization, and odorant sensory neuron localization. *Genomics*, 2002. 80(3): p. 344.
7. JM, A., et al., Regulated expression of ATF5 is required for the progression of neural progenitor cells to neurons. *Journal of Neuroscience the Official Journal of the Society for Neuroscience*, 2013. 23(11): p. 4590.
8. Britt, W.J., et al., Cell surface expression of human cytomegalovirus (HCMV) gp55-116 (gB): use of HCMV-recombinant vaccinia virus-infected cells in analysis of the human neutralizing antibody response. *Journal of Virology*, 1990. 64(3): p. 1079.
9. Weber, B., et al., Humoral immune response to human cytomegalovirus infection: diagnostic potential of immunoglobulin class and IgG subclass antibody response to human cytomegalovirus early and late antigens. *Clinical Investigator*, 1993. 71(4): p. 270-276.
10. Van, Z.J., et al., Humoral immune response against human cytomegalovirus (HCMV)-specific proteins after HCMV infection in lung transplantation as detected with recombinant and naturally occurring proteins. *Clin Diagn Lab Immunol*, 1995. 2(2): p. 214-218.
11. Hsu, C.H., et al., HCMV IE2-mediated inhibition of HAT activity downregulates p53 function. *Embo Journal*, 2004. 23(11): p. 2269-2280.
12. Braendstrup, P., et al., Identification and HLA-Tetramer-Validation of Human CD4+ and CD8+ T Cell Responses against HCMV Proteins IE1 and IE2. *Plos One*, 2014. 9(4): p. e94892.
13. Stinski, M.F. and D.T. Petrik, Functional Roles of the Human Cytomegalovirus Essential IE86 Protein. *Curr Top Microbiol Immunol*, 2008. 325(1): p. 133-152.
14. Lang, D., et al., Functional interaction between the human cytomegalovirus 86-kilodalton IE2 protein and the cellular transcription factor CREB. *Journal of Virology*, 1995. 69(10): p. 6030.
15. Mason, J.L., et al., ATF5 regulates the proliferation and differentiation of oligodendrocytes. *Molecular & Cellular Neuroscience*, 2005. 29(3): p. 372.
16. Zhou, D., et al., Phosphorylation of eIF2 Directs ATF5 Translational Control in Response to Diverse Stress Conditions. *Journal of Biological Chemistry*, 2008. 283(11): p. 7064-7073.
17. JM, A., et al., Selective destruction of glioblastoma cells by interference with the activity or expression of ATF5. *Oncogene*, 2006. 25(6): p. 907-16.
18. Hu, M., et al., Human cytomegalovirus immediate-early protein promotes survival of glioma cells through interacting and acetylating ATF5. *Oncotarget*, 2017. 8(19): p. 32157.
19. Huang, R., et al., Association between human cytomegalovirus infection and histone acetylation level in various histological types of glioma. *Oncology Letters*, 2015. 10(5): p. 2812.
20. Rodems, S.M., C.L. Clark, and D.H. Spector, Separate DNA Elements Containing ATF/CREB and IE86 Binding Sites Differentially Regulate the Human Cytomegalovirus UL112-113 Promoter at Early and Late Times in the Infection. *Journal of Virology*, 1996. 70(5): p. 2697-2707.